one of the burrows the previous day. Subsequently, Mr. Carlisle reported (April 30th, 1966) that there is still a pair of wombats living in the burrows and that the main colony has been located a few miles away.

The burrows appear to be typical of wombats, with a diameter of about 2 feet and deeply scratched walls. Fresh tracks were seen, and fresh faeces were collected. These faeces were lodged with the Western Australian Museum and were examined by Mr. P. Aitken of the South Australian Museum who agreed that they were most probably *Lasiorhinus* (pers. comm. to Mr. D. Merrilees).

The burrows occur on the western side of a clay pan which appears as a dark patch 3.18 inches on a bearing of 51.5° from the centre point of air photo Culver Run 2, CAF4009-5725 (1961 photography), approximately 125° 25′ E., 126° 10′ S.

Another wombat burrow about 3 miles to the south-west of the first site was in use when discovered by Mr. Carlisle in 1965, but it also was abandoned when he showed it to us in January, 1966.

—DAVID C. LOWRY, Geological Survey, Perth.

Further Notes on the W.A. Pitcher Plant.—Since publication of my paper in *W. Aust. Nat.*, 10 (1) 1965: 1-7, the need for several adjustments has arisen. These amendments and additions are set out hereunder:

3rd paragr. from bottom (line 3)—insert inverted commas before *Cephalotus.*

Page 6 (lines 3 and 4)—

Mrs. Rica Erickson, 1 Boronia Ave., Nedlands, has kindly drawn my attention to a startling discrepancy between Hooker's printed quotation from James Drummond and the actual words in his original letter (21 Feb., 1844), of which she has a copy. Drummond did *not* write "Tomorrow I start for Freemantle, where *Cephalotus follicularis* grows," but used the following words:

> "I observe that the passengers for London by the 'Shepherd' are warned to be on board on Saturday next, but a delay of a day or two may occur, and I shall set out tomorrow for Fremantle and endeavour to get on board some boxes of *Cephalotus follicularis:* one box I will send for your garden."

It is hardly credible that the erudite and scrupulously honest William Hooker would misquote deliberately, but in this case his very poor editing remains inexcusable. Mrs. Erickson has come across other instances of present-day confusion resulting from faulty references by Hooker to Drummond's collections and statements.

Page 7 (lines 13 and 14)—

Between the lines beginning LLOYD and PELLOE, insert an extra reference:

MACFARLANE, J. M. 1911. Cephalotaceae. *Pflanzenreich IV* 116 (Heft 47): 1-15, fig. 1-4.

This was an inadvertent omission of important literature, for which the writer must accept full blame. Incidentally Macfarlane (*l.c.* p. 13, line 14), alluding to the Geographical Distribution of *Cephalotus,* repeats the myth "Its discoverer Labillardiere found it at Esperance Bay."

—J. H. WILLIS, National Herbarium, Melbourne.

Preservation Techniques for Algae

1. Field collections: Problems of carrying newspaper and presses, washing and arranging specimens, and changing of pressing papers can be obviated by using a drip-wet method. Algae are collected and steeped in the standard algal preservative of 6% formalin in seawater for at least 3-4 hours. The material is drained of solution and immediately placed in waterproof containers. Plastic bags, with wire ties, are recommended for ease of transport, convenience, and easy marking with a felt marker pen. In this condition material can be stored for up to 4 months. In the convenience of the laboratory the material may be sorted and conventional herbarium mounts prepared. Since the algae have been soaked in formalin solution fungal growth on the herbarium sheets is prevented. If desired, the wet material can be hand sectioned for anatomical details or reproductive structures.

2. A mountant for slides of algae: Glycerine jelly has long been a standard mountant for algae, but never completely satisfactory as a hard, permanent preparation. A pure corn syrup, designed for cooking purposes, has proven satisfactory for permanent mounts.

The algal material is fixed (in 6% formalin, or other appropriate fixative) and then either mounted directly or stained if necessary before mounting. The corn syrup is diluted to a 1 : 1 solution with distilled water (a fungicide such as phenol must be added as this sugar solution is ideal for fungal growth). A drop of solution is placed on a slide with the algal specimen, allowed to harden for a few minutes, and then a cover slip is added. Overnight the syrup will harden into a permanent mount. Many other plant tissues can also be mounted by this means.

Gentian violet is a useful algal stain and, made up in aqueous solution, can be used to pre-stain the syrup mountant solution. This simplifies procedures as the material may be stained and mounted simultaneously.

The corn syrup is manufactured under the trade name of KARO, by the Best Foods Division, Corn Products Company, New York. It is available in 16 fluid ounce bottles from many food and gourmet counters.

—B. M. ALLENDER, Department of Botany, University of Western Australia.